Parasitoids associated with South American fruit fly in native fruits in the state of Santa Catarina, Brazil

Janaína Pereira dos Santos¹, Alexandre Carlos Menezes-Netto² and Juracy Caldeira Lins Junior³

Abstract - In Southern of Brazil, the South American fruit fly (Anastrepha fraterculus) is the main insect pest of native and exotic fruit trees. This study was conducted in agricultural areas of seven municipalities belonging to the Alto Vale do Rio do Peixe region, in the state of Santa Catarina, Brazil, from 2015 to 2022. It aimed to evaluate the entomofauna of microhymenopteran parasitoids associated with A. fraterculus in native fruit trees. A sample of 200 fruits of Eugenia involucrata (cherry of the Rio Grande), Eugenia pyriformis ('uvaia'), Acca sellowiana ('feijoa'), Psidium cattleyanum (yellow and red cattley guava), Campomanesia xanthocarpa ('guabiroba') and Campomanesia guazumifolia ('sete capotes') per municipality and in each crop season was collected to determine the parasitism index and perform the faunistic analysis of parasitoid species. Seven parasitoid species were recorded, three from the family Figitidae and four from the family Braconidae. Aganaspis pelleranoi was the most often recorded species, with a constant presence, very abundant, and dominant incidence in the investigated region. Parasitism rates ranged from 0.2% to 39%, emphasizing the need for conservation-oriented and applied biological control studies.

Index terms: Anastrepha fraterculus; Natural biological control; Micro-hymenopteran.

Parasitoides associados à mosca-das-frutas sul-americana em frutíferas nativas em Santa Catarina, Brasil

Resumo - No Sul do Brasil, a mosca-das-frutas sul-americana (*Anastrepha fraterculus*) é o principal inseto-praga das frutíferas nativas e exóticas. O estudo foi realizado em áreas agrícolas de sete municípios pertencentes à região do Alto Vale do Rio do Peixe, em Santa Catarina, Brasil, no período de 2015 a 2022. Este estudo teve como objetivo avaliar a entomofauna de microhimenópteros parasitoides associados à *A. fraterculus* em frutíferas nativas. Uma amostra de 200 frutos de *Eugenia involucrata* (cerejeira-do-mato), *Eugenia pyriformis* (uvaieira), *Acca sellowiana* ('feijoa'), *Psidium cattleianum* (araçazeiros amarelo e vermelho), *Campomanesia xanthocarpa* (guabirobeira) e *Campomanesia guazumifolia* (sete-capotes) foram coletadas por município e em cada safra agrícola para determinar os índices de parasitismo e a análise faunística das espécies de parasitoides. Foram registradas sete espécies de parasitoides, três da família Figitidae e quatro de Braconidae. A espécie mais frequente foi *Aganaspis pelleranoi*, apresentando ocorrência constante, muito abundante e dominante na região. Os índices de parasitismo variaram entre 0,2 a 39%, reforçando a necessidade de estudos sobre controle biológico, por conservação e/ou aplicado.

Termos para indexação: Anastrepha fraterculus; Controle biológico natural; Micro-hymenoptera.

Introduction

The South American fruit fly, *Anastrepha fraterculus* (Wiedemann, 1830) (Diptera: Tephritidae), is one of the main pests affecting native and

exotic temperate climate fruit trees in Southern Brazil (GARCIA & NORRBOM, 2011; ROSA et al., 2017; SANTOS & GUIMARÃES, 2018). The application of toxic baits and insecticides in the cover area are the main practices used by fruit

growers to suppress fruit fly populations. However, the use of insecticides is increasingly limited due to the demand of importing countries for fresh fruits without chemical residues (BORGES et al., 2015). In addition, Brazilian

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consumers and producers are more environmentally conscious; therefore, management practices applied to *A. fraterculus* need to improved and other control strategies must be adopted (DIAS et al., 2018). In this sense, biological control can play a crucial role in the fruit sector (NAVA, 2007).

Biological control is a promising alternative for managing Tephritidae fruit flies in Brazil, mainly by implementing of native parasitoids (DIAS et al., 2022). One of the management strategies used is the maintenance of refuges adjacent to orchards, based on cultural control and the use of selective insecticides (NAVA, 2007). These practices alow for the establishment and the reproduction of parasitoid species found in agroecosystems (SANTOS, 2022).

Several native fruit tree species cultivated in Southern Brazil are host to flies of the genus Anastrepha, mainly those of the family Myrtaceae (GARCIA & NORRBOM, 2011; SANTOS et al., 2018; BOLDO et al., 2019; SANTOS et al., 2022). It is essential to detect and measure fruit fly populations in these hosts to better understand their infestation and parasitism levels (SANTOS et al., 2018; BOLDO et al., 2019). These steps play an essential role in the implementation of biological control techniques (SANTOS et al., 2018). The last surveys on parasitoids associated with A. fraterculus in the state of Santa Catarina were carried out in the 1990s (LEONEL JUNIOR et al., 1995; GUIMARÃES et al., 2000; NORA et al., 2000; GARCIA & CORSEUIL, 2004), highlighting the need to conduct new studies on this topic. Thus, this study aimed to evaluate the entomofauna of micro-hymenopteran parasitoids associated with the South American fruit fly, A. fraterculus, in native fruit trees in the Alto Vale do Rio do Peixe region (AVRP), state of Santa Catarina, Brazil.

Materials and Methods

This study was carried out from 2015 to 2022, and the harvesting season was from October to April of each year, which corresponds to the period of fruit ripening of native species cultivated in the AVRP region, state of Santa Catarina. The following native fruit trees were assessed: Eugenia involucrata DC. (cherry of the Rio Grande), Eugenia pyriformis Cambess. ('uvaia'), Acca sellowiana (O. Berg.) Burret ('feijoa'); Psidium cattleianum Sabine (red and yellow cattley guava), Campomanesia xanthocarpa O. Berg. ('guabiroba'); Campomanesia guazumifolia (Cambess.) O. Berg. ('sete capotes') (Table 1). Fruit sampling was carried out in the following municipalities: Cacador; Calmon; Macieira; Matos Costa; Rio das Antas; Videira and Lebon Régis. In Cacador, sampling was carried out from 2015 to 2019; in Calmon, Macieira, Matos Costa, Rio das Antas and Videira, sampling was carried out from 2019 to 2022, and in Lebon Régis, sampling was conducted from 2020 to 2022 (Table 1).

Samples of 200 ripe fruits of each species per municipality were collected in each crop season. The fruits were collected randomly both from the ground and treetops. These plants were not sprayed with pesticides and were located near commercial orchards where Malus domestica Borkh. (apple), Pyrus spp. (pear), Prunus persica L. Batsch (peach), Prunus domestica L. (plum) and Vitis spp. (grape) are grown. Fruits sampled from the ground and treetops were categorized as single samples and assessed together because they were at the same stage of the ripening.

The native fruit species studied had an intense and short ripening period of about 15 to 20 days. Thus, the samples collected here only included fruits that were fully ripe, i.e., that were most susceptible to fruit fly infestation. The analysis of fruit ripening stage included the following parameters: pulp firmness, measured with a digital motorized penetrometer equipped with a 3mm diameter plunger, and harvest time of each native fruit tree species, established based on data reported in previous surveys conducted in the AVRP region. Changes in fruit epidermis color were also monitored.

The fruits were placed in plastic containers covered with a layer of sterilized sand of approximately 1cm and kept in a room with controlled climatic conditions (25 ± 1°C; 60 ± 10% relative humidity; 12h photophase) at the Entomology Laboratory of the Empresa de Pesquisa Agropecuária e Extensão Rural de Santa Catarina (Epagri), at the Experimental Station of Caçador "José Oscar Kurtz". After seven, 14 and 21 days of incubation, the sand was examined for puparia counting. Puparia were kept in Petri dishes covered with sterilized sand (substrate) and placed in cages (40.0 x 29.0 x 51.5cm) or in 48 well cell culture plates to favor the emergence of adults.

The emerged fruit fly specimens were stored in plastic vials (50mL) filled with 70% alcohol. They were then identified to the species level according to Zucchi (2000). Parasitoid specimens were identified using specific dichotomous keys described by Canal & Zucchi (2000) and Guimarães et al. (2000).

The parasitism index was determined using the following formula: PI% = (P TA⁻¹) x 100, in which: P = total number of emerged parasitoids and TA = total number of emerged adults (fruit flies + parasitoids). The relative frequency of species, expressed in percentage (%), was calculated by considering the proportion of individuals belonging to the same species and the total number of individuals in the sample.

Faunistic analysis was carried out in

Table 1. Period of fruit collection of native fruit tree species in different municipalities of the Alto Vale do Rio do Peixe region, state of Santa Catarina, Brazil (crop seasons from 2015/2016 to 2021/2022)

Tabela 1. Período de coleta de frutos de frutíferas nativas em diferentes municípios da região do Alto Vale do Rio do Peixe, Santa Catarina, Brasil (safras 2015/2016 a 2021/2022)

Caçador Oct/2015 Nov/2016 Nov/2017 Nov/2018 - - - Macieira - - - Nov/2019 Oct/2020 Nov/2021 Videira - - - Nov/2020 Nov/2021 Campomanesia xanthocarpa ('guabiroba') - - - - Caçador Dec/2015 Dec/2016 Dec/2017 Dec/2018 - - - Rio das Antas - - - - Dec/2019 Dec/2020 Dec/2021 Macieira - - - - Dec/2019 Dec/2020 Dec/2021 Matos Costa - - - - Dec/2021 Dec/2020 Dec/2021 Calmon - - - - Dec/2021 Dec/2020 Dec/2021 Videira - - - - - Dec/2020 Dec/2021 Campomanesia guazumifolia ('sete capotes') - - - Feb/2020	Native fruit trees species Municipalities	Crops/fruit collection months							
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Carpador annihocarpa ('guabiroba') Caçador Dec/2015 Dec/2016 Dec/2017 Dec/2018	Macieira	-	-	-	-	Nov/2019	Oct/2020	Nov/2021	
Caçador Dec/2015 Dec/2016 Dec/2017 Dec/2018 -	Videira	-	-	-	-	-	Nov/2020	Nov/2021	
Rio das Antas	Campomanesia xanthocarpa ('guabiroba')								
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	Matos Costa	-	-	-	-	Mar/2020	Mar/2021	Mar/2022	
Lebon Régis Mar/2021 -	Videira	-	-	-	-	Mar/2020	Feb/2021	Feb/2022	
	Lebon Régis	-	-	-	-	-	Mar/2021	-	

Dashes (-) represent the crop seasons when no fruits were collected.

The species were divided into the following frequency classes: little frequent (LF) - frequency below the lower limit of the confidence interval (CI) of the mean; frequent (F) frequency ranging between the lower and upper limits of the CI of the mean; very frequent (VF) - frequency above the upper limit of the CI of the mean.

Constancy rates were classified as follows: constant (W) - higher than the CI limit; accessory (Y) - within the CI;

and accidental (Z) - below the lower CI limit.

For dominance, species were classified as follows: super dominant (SD) and dominant (D) - frequency above the dominance limit; and

non-dominant (ND) - frequency below than the dominance limit.

Abundance was divided into the following classes: rare (r) - number of individuals below the lower limit of the CI of the mean; dispersed (d) - number of individuals between the lower and upper limits of the CI of the mean; common (c) - number of individuals between the lower and upper limits of the CI of the mean; abundant (a) - number of individuals meeting the upper limits of the CI of the mean; and very abundant (va) - number of individuals above the upper limit of the CI of the mean.

The researchers responsible for the current study were duly authorized by the Chico Mendes Institute for Biodiversity Conservation (ICMBio) and Authorization and Information System in Biodiversity (SISBIO).

Results and Discussion

total, 23,709 puparia of Anastrepha spp. were observed in this study, of which 7,414 flies of the species A. fraterculus emerged. In 355 of these puparia parasitoids emerged. The remaining pupae were unviable. The highest abundance rate of A. fraterculus observed among tephritids in the state of Santa Catarina had already been recorded in citrus (GARCIA & LARA, 2006), peach (ALBERTI et al., 2009; GARCIA & NORRBOM, 2011), passion fruit (ALBERTI et al., 2009), pear (ROSA et al., 2017), plum and 'feijoa' (GARCIA & NORRBOM, 2011; ROSA et al., 2017), and apple (SANTOS & GUIMARÃES, 2018).

Seven parasitoid species were recorded, four of which belonged to the family Braconidae (Opiinae): Doryctobracon brasiliensis (Szépligeti, 1911); Doryctobracon areolatus (Szépligeti, 1911); Opius sp. and Utetes anastrephae (Viereck, 1913); while

three belonged to the family Figitidae Aganaspis (Eucoilinae): pelleranoi (Brèthes, 1924); Aganaspis nordlanderi Wharton, 1998 and Odontosema anastrephae Borgmeier, 1935 (Table 2). The subfamilies Eucoilinae (Figitidae) and Opiinae (Braconidae) included the most representative species used for biological control of tephritids (CANAL & ZUCCHI, 2000; GUIMARÃES et al., 2003; DIAS et al., 2022). The main genus of Anastrepha parasitoids observed in the state of Santa Catarina belonged to the families Braconidae, Diapriidae and Figitidae (GARCIA & COURSEUIL, 2004; SANTOS & GUIMARÃES, 2018). Some of the Braconidae species we identified in this work had previously been recorded in association with Anastrepha in the state of Santa Catarina, such as: U. anastrephae (LEONEL JUNIOR et al., 1995; NORA et al., 1995; NORA et al., 2000; GARCIA & COURSEUIL, 2004); D. areolatus, D. brasiliensis (LEONEL JUNIOR et al., 1995; NORA et al., 1995; NORA et al., 2000; GARCIA & COURSEUIL, 2004; SANTOS & GUIMARÃES, 2018), and Opius bellus Gahan, 1930 (LEONEL JUNIOR et al., 1995; NORA et al., 1995; NORA et al., 2000; GARCIA & COURSEUIL, 2004). According to Canal & Zucchi (2000), U. anastrephae was misidentified as Opius tomopagliae (Costa Lima, 1937). As a resulted, this species currently lacks records in the state of Santa Catarina.

The following Figitidae species (Eucoilinae) recorded in this study had already been reported in the state of Santa Catarina: *O. anastrephae* (GUIMARÃES et al., 2000; GARCIA & COURSEUIL, 2004); *A. pelleranoi* (GUIMARÃES et al., 2000; NORA et al., 2000; GARCIA & COURSEUIL, 2004; SANTOS & GUIMARÃES, 2018); and *A. nordlanderi* (SANTOS & GUIMARÃES, 2018). Although Garcia and Courseuil (2004) reported *Trichopria anastrephae* Lima, 1940 (Diapriidae) in fruits of *Psidium quajava* L. (guava), this species

was not recorded in the present study, probably because it was a not evaluated host

Of the all parasitoid species recorded in this study, only *A. nordlanderi* had not been recorded in surveys carried out in the state of Santa Catarina in the 1990s. In fact, *A. nordlanderi* was first documented in Southern Brazil in 2015, specifically in Caçador, in the state of Santa Catarina (SANTOS & GUIMARÃES, 2018). In our study, this species was recorded in the municipalities of Caçador and Macieira (Table 2).

Red cattley guava and cherry of the Rio Grande were the hosts harboring the highest number of parasitoid species, comprising six of the seven species recorded here (Table 2). These findings corroborate those reported by Garcia and Corseuil (2004), who found that cattley guava, cherry of the Rio Grande and guava were the hosts harboring the highest number of parasitoid species in A. fraterculus in Western Santa Catarina. Nora et al. (1995) recorded parasitoids associated with A. fraterculus in fruits of 'feijoa' (D. areolatus, D. brasiliensis, bellus and U. anastrephae), 'guabiroba' (D. brasiliensis, O. bellus and U. anastrephae), 'sete capotes' (D. areolatus and D. brasiliensis) and yellow cattley guava (D. brasiliensis) in the AVRP region. In the study mentioned above, Eucoilinae (not identified at the species level) was also recorded in fruits of all hosts assessed.

Parasitism rates ranged from 0.2% to 39%, with variations between fruit trees and collection municipalities (Table 2). According to Nora et al. (2000), the level of parasitism in the state of Santa Catarina is variable and influenced by the host plant species in which fruit flies develop. According to Canal and Zucchi (2000), the natural parasitism of fruit flies varies depending on aspects such as host fruit, location and time of collection. However, these rates rarely exceed 50%, which is considered low

Table 2. Parasitoids associated with Anastrepha fraterculus (Diptera: Tephritidae) and parasitism index recorded for native fruit tree species, in the Alto Vale do Rio do Peixe region, state of Santa Catarina, Brazil (crop seasons from 2015/2016 to 2021/2022)

Tabela 2. Parasitoides associados à Anastrepha fraterculus (Diptera: Tephritidae) e índice de parasitismo em frutos de frutíferas nativas, na região do Alto Vale do Rio do Peixe, Santa Catarina, Brasil (safras 2015/2016 a 2021/2022)

Native fruit trees species	Parasitoid Family/Species	Collection Municipality (Parasitism index %)			
Daiding antidaing an	Figitidae Aganaspis pelleranoi Aganaspis nordlanderi	Caçador (0.2); Rio das Antas (7.2); Videira (4.7); Lebor Régis (1.5) Caçador (0.2)			
Psidium cattleianum (red cattley guava)	Braconidae Doryctobracon brasiliensis Doryctobracon areolatus	Caçador (7); Videira (1.4); Matos Costa (0.3); Rio das Antas (3); Lebon Régis (2) Caçador (4.4); Videira (16.3); Matos Costa (0.3); Lebon Régis (3)			
	Utetes anastrephae Opius sp.	Videira (1.5) Rio das Antas (0.4); Videira (4.5)			
Psidium cattleianum (yellow cattley guava)	Figitidae Aganaspis pelleranoi Aganaspis nordlanderi Odontosema anastrephae	Caçador (2.5); Macieira (23); Rio das Antas (0.8); Videira (2.4) Macieira (9) Rio das Antas (0.9)			
	Braconidae Doryctobracon brasiliensis	Caçador (12); Matos Costa (11.5); Macieira (5)			
	Doryctobracon areolatus	Macieira (15); Rio das Antas (2)			
Acca sellowiana	Figitidae Aganaspis pelleranoi	Caçador (4.2)			
('feijoa')	Braconidae Doryctobracon brasiliensis	Caçador (4); Videira (1.3)			
	Doryctobracon areolatus	Caçador (4.5); Videira (3)			
Eugenia involucrata (cherry of the Rio Grande)	Figitidae Aganaspis pelleranoi	Caçador (4); Videira (3.3)			
	Aganaspis nordlanderi Odontosema anastrephae	Caçador (0.9) Videira (0.6)			
	Braconidae Doryctobracon brasiliensis Doryctobracon areolatus Utetes anastrephae	Macieira (3.3); Videira (0.8) Caçador (5); Macieira (1.2) Caçador (7); Macieira (8); Videira (10)			
Campomanesia xanthocarpa ('guabiroba')	Figitidae Aganaspis pelleranoi Aganaspis nordlanderi	Videira (31); Matos Costa (9); Lebon Régis (1) Caçador (6)			
	Braconidae Doryctobracon brasiliensis	Calmon (4); Videira (7)			
	Utetes anastrephae	Videira (2); Macieira (6); Rio das Antas (39); Lebon Régis (1)			
Eugenia pyriformis ('uvaia')	Braconidae Doryctobracon areolatus	Caçador (6)			
Campomanesia guazumifolia ('sete capotes')	Figitidae Aganaspis pelleranoi	Videira (0.9)			

for integrated pest management programs. According to the authors, natural parasitism can be enhanced by conservation-oriented or applied biological control based on massive releases. According to Nava (2007), several potential species have already been listed for the applied biological control of *A. fraterculus* in the state of Rio Grande do Sul, Brazil. Among them, *Opius* sp., *D. brasiliensis*, *D. areolatus* and *A. pelleranoi* were found to parasitize up to 40% of fruit fly larvae in different orchards.

Of the seven species studied here, A. pelleranoi was not associated with 'uvaia'. Therefore, this parasitoid species was recorded in most assessed hosts (Table 2). It was also the most frequent species, accounting for 31.8% of the emerged parasitoids, and was constant, very abundant and dominant in the AVRP region (Table 3). Aganaspis pelleranoi was not recorded in Calmon. Moreover, it was a little frequent and had an accidental, dispersed and nondominant incidence in Macieira. On the other hand, in the other municipalities, it had a frequent or very frequent incidence, was mostly dominant (Table 3). According to Garcia & Corseuil (2004), A. pelleranoi was the most frequent species recorded in Western Santa Catarina (25.6%), followed by D. brasiliensis (21.1%) and D. areolatus (18.6%). These authors observed that cattley guava was a common host for these three parasitoid species, which is in line with the present results.

Utetes anastrephae was the second most frequent species. It was associated with three hosts (Table 2) and accounted for 24% of emerged parasitoids. It was very frequent and had an accessory, abundant and dominant incidence in the AVRP region (Table 3). The species D. areolatus and D. brasiliensis were associated with five hosts (Table 2) accounting for 18.0% and 21.7% of

emerged parasitoids, respectively, and showing frequent, common and dominant incidence in the AVRP region. Odontosema anastrephae (1.1%), A. nordlanderi (1.4%), and Opius sp. (2.0%) were the least frequent species, with an accidental incidence in the AVRP region. Opius sp. and O. anastrephae were observed only in the cities of Videira and Rio das Antas, while A. nordlanderi was recorded in Macieira and Caçador (Table 2). A. nordlanderi and O. anastrephae had a rare and non-dominant incidence, while Opius sp. had a dispersed and dominant incidence (Table 3). Calmon only recorded one parasitoid of the species D. brasiliensis: therefore, it was not possible to perform its faunistic analysis.

Since the use of insecticides is increasingly limited, especially in fruits grown for export, it is necessary to find other strategies focused on controlling fruit flies in orchards based on integrated pest management. Thus, studies focused on investigating the diversity of parasitoid species can help the biological control of fruit flies by providing essential information on fruit species that act as hosts for these natural enemies, besides encouraging future studies aimed at applied biological control.

Conclusion

- The parasitoids of the species A. pelleranoi, A. nordlanderi, O. anastrephae (Figitidae), D. brasiliensis, D. areolatus, Opius sp. and U. anastrephae (Braconidae) were associated with A. fraterculus in fruits from native host trees grown in the Alto Vale do Rio Peixe region, state of Santa Catarina;
- Aganaspis pelleranoi was the most frequent species, with a constant, very abundant and dominant incidence;
 - The region showed a diversity of

parasitoid species, although at low parasitism rates, and it is necessary to conduct new studies focused on investigating conservation-oriented and applied biological control processes.

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References

ALBERTI, S.; GARCIA, F.R.M.; BOGUS, G.M. Moscas-das-frutas em pomares de pessegueiro e maracujazeiro, no município de Iraceminha, Santa Catarina, Brasil. **Ciência Rural**, Santa Maria, v.39, n.5, p.1565-1568, 2009.

BOLDO, R.S.; KOVALESKI, A.; ROSA, J.M.; BOFF, M.I.C.; FRANCO, C.R. Development of *Anastrepha fraterculus* (Wiedemann, 1830) (Diptera: Tephritidae) in different host fruits. **Journal of Agricultural Science,** Otawa, v.11, n.8, p.273-279, 2019.

BORGES, R.; MACHOTA JUNIOR, R.; BOFF, M.I.C.; BOTTON, M. Efeito de iscas tóxicas sobre *Anastrepha fraterculus* (Wiedemann) (Diptera: Tephritidae). **BioAssay**, Piracicaba, v.10, n.3, p.1-8, 2015.

CANAL, N.A.; ZUCCHI, R.A. Parasitóides – Braconidae. *In:* MALAVASI, A.; ZUCCHI, R.A. (orgs.). **Moscas-das-frutas de importância econômica no Brasil:** conhecimento básico e aplicado. Ribeirão Preto: Holos, 2000. p.119-126.

Table 3. Faunistic analysis applied to parasitoid species associated with Anastrepha fraterculus (Diptera: Tephritidae) in the Alto Vale do Rio do Peixe region, Santa state of Catarina, Brazil (crop seasons from 2015/2016 to 2021/2022)

Tabela 3. Análise faunística das espécies de parasitoides associadas à Anastrepha fraterculus (Diptera: Tephritidae) na região do Alto Vale do Rio do Peixe, Santa Catarina, Brasil (safras 2015/2016 a 2021/2022)

Collection	Parasitoid Family/Species	Relative	Classes				
Municipality	r arasitora r anniy/ species	frequency (%)	Frequency ¹	Constancy ²	Abundance ³	Dominance ⁴	
	Figitidae						
Caçador	Aganaspis pelleranoi	32.3	F	Υ	С	D	
	Aganaspis nordlanderi	4.2	LF	Z	d	ND	
	Braconidae						
	Doryctobracon areolatus	15.6	F	Z	С	D	
zaçadoi	Doryctobracon brasiliensis	40.6	VF	Z	va	D	
	Utetes anastrephae	7.3	F	Z	С	D	
	Figitidae						
	Aganaspis pelleranoi	38.9	VF	Υ	va	D	
	Odontossema anastrephae	2.8	LF	Z	r	ND	
	Braconidae						
	Doryctobracon areolatus	8.3	F	Z	С	ND	
Rio das Antas	Doryctobracon brasiliensis	13.9	F	Z	С	ND	
	Utetes anastrephae	19.4	F	Z	С	D	
	Opius sp.	16.7	F	Υ	С	D	
	Figitidae						
	Aganaspis pelleranoi	36.7	VF	Υ	va	D	
	Odontossema anastrephae	1.8	LF	Z	d	ND	
	Braconidae						
	Doryctobracon areolatus	18.3	F	Z	С	D	
/ideira	Doryctobracon brasiliensis	7.7	F	Υ	С	D	
	Utetes anastrephae	34.9	VF	Z	va	D	
	Opius sp.	0.6	LF	Z	d	ND	
	Figitidae				-		
	Aganaspis pelleranoi	8.0	LF	Z	d	ND	
	Aganaspis nordlanderi	2.6	LF	Z	r	ND	
	Braconidae						
	Doryctobracon areolatus	31.6	F	Υ	С	D	
Macieira	Doryctobracon brasiliensis	28.9	F	Y	С	D	
	Utetes anastrephae	28.9	F	Z	С	D	
	Figitidae						
	Aganaspis pelleranoi	11.1	F	Υ	va	ND	
	Braconidae		· · · · · · · · · · · · · · · · · · ·			.,	
Matos Costa	Doryctobracon areolatus	11.1	F	Υ	va	ND	
	Doryctobracon brasiliensis	77.8	VF	W	va	D	
	Figitidae	77.10					
	Aganaspis pelleranoi	33.3	F	Υ	С	ND	
	Braconidae	33.3	•			112	
	Doryctobracon areolatus	33.3	F	Z	С	ND	
Lebon Régis	Doryctobracon brasiliensis	16.7	F	Z	С	ND	
	Utetes anastrephae	16.7	F	Z	С	ND	
	Figitidae	10.7			· ·	ND	
	Aganaspis pelleranoi	31.8	VF	W	va	D	
	Aganaspis nordlanderi	1.4	LF	Z	r	ND	
	Odontossema anastrephae	1.1	LF	Z	r	ND	
Alto Vale do Rio do Peixe region	Braconidae	1.1	LF			ND	
	Doryctobracon areolatus	18.0	F	Υ	С	D	
	Doryctobracon brasiliensis	21.7	F	W		D	
					С		
	Utetes anastrephae	24.0	VF	Y	a	D	
	Opius sp.	2.0	LF	Z	d	D	

^{1 -} Frequency: LF = little frequent; F = frequent; VF= very frequent.

^{2 -} Constancy: Z = accidental; Y = accessory; W = constant.

^{3 -} Abundance: r = rare; d = dispersed; c = common; va = very abundant.

^{4 -} Dominance: ND = non-dominant; D = dominant.

DIAS, N.P.; ZOTTI, M.J.; MONTOYA, P.; CARVALHO, I.R.; NAVA, D.E. Fruit fly management research: a systematic review of monitoring and control tactics in the world. **Crop Protection**, Guildford, v.112, p.187-200, 2018.

DIAS, N.P.; MONTOYA, P.; NAVA, D.E. A 30-year systematic review reveals success in tephritid fruit fly biological control research. **Entomologia Experimentalis et Applicata,** Dordrecht, n.170, p.370-384, 2022.

GARCIA, F.R.M.; CORSEUIL, E. Native hymenopteran parasitoids associated with fruit flies (Diptera: Tephritidae) in Santa Catarina State, Brazil. **Florida Entomologist,** Gainesville, v.87, n.4, p.517-521, 2004.

GARCIA, F.R.M.; LARA, D.B. de. Análise faunística e flutuação populacional de mosca-das-frutas (Diptera, Tephritidae) em pomar cítrico no município de Dionísio Cerqueira, Santa Catarina. **Biotemas,** Florianópolis, v.19, n.3, p.65-70, 2006.

GARCIA, F.R.M.; NORRBOM, A.L. Tephritoid flies (Diptera, Tephritoidea) and their plant hosts from state of Santa Catarina in Southern Brazil. **Florida Entomologist,** Gainesville, v.94, n.2, p.151-157, 2011.

GUIMARÃES, J.A.; DIAZ, N.B.; ZUCCHI, R.A. Parasitóides - (Figitidae: Eucoilinae). *In:* MALAVASI, A.; ZUCCHI, R.A. (orgs.). **Moscas-das-frutas de importância econômica no Brasil:** conhecimento básico e aplicado. Ribeirão Preto: Holos, 2000. p.127-134.

GUIMARÃES, J.A.; GALLARDO, F.E.; DIAZ, N.B.; ZUCCHI, R.A. Eucoilinae species (Hymenoptera: Cynipoidea: Figitidae) parasitoids of fruit-infesting dipterous in Brazil: identity, geographical distribution and host associations. **Zootaxa**, v.278,

p.1-23, 2003.

LEONEL JR., F.L.; ZUCCHI, R.A.; WHARTON, R.A. Distribution and tephritid hosts (Diptera) of braconid parasitoids (Hymenoptera) in Brazil. International Journal of Pest Management, London, v.41, n.4, p.208-213, 1995.

MORAES, R.C.B.; HADDAD, M.L.; SILVEIRA NETO, S.; REYES, A.E.L. Software para análise faunística. *In:* SIMPÓSIO DE CONTROLE BIOLÓGICO, 8., 2003, São Pedro. **Anais do 8º Siconbiol[...].** 2003. v.1, n.1, p.195.

NAVA, D.E. Controle biológico de insetos-praga em frutíferas de clima temperado: uma opção viável, mas desafiadora. Embrapa: Pelotas, 2007. 20p. (Embrapa Clima Temperado. Documentos, 208).

NORA, I.; CANAL, N.A.; ZUCCHI, R.A.; BRANCO, E.S. Levantamento de parasitóides (Hym., Braconidae) de *Anastrepha* (Dip., Tephritidae) na região do Alto Vale do Rio do Peixe, SC. *In*: CONGRESSO BRASILEIRO DE ENTOMOLOGIA, 15., ENCONTRO NACIONAL DE FITOSSANITARISTAS, 6., SIMPÓSIO INTEGRADO DE MANEJO DE PRAGAS, 2., 1995, Caxambu. **Resumos[...].** Lavras: ESAL, 1995. p.298.

NORA, I.; HICKEL, E.R.; PRANDO, H.F. Moscas-das-frutas nos estados brasileiros: Santa Catarina. *In:* MALAVASI, A.; ZUCCHI, R. A. (orgs). **Moscas-das-frutas de importância econômica no Brasil:** conhecimento básico e aplicado. Ribeirão Preto: Holos. 2000. p.271-276.

ROSA, J.M.; ARIOLI, C.J.; SANTOS, J.P.; MENEZES-NETTO, A.C.; BOTTON, M. Evaluation of food lures for capture and monitoring of *Anastrepha fraterculus* (Diptera: Tephritidae) on temperate fruit

trees. **Journal of Economic Entomology,** Laham, v.110, p. 995-1001, 2017.

SANTOS, J.P.; GUIMARÃES, J.A. Parasitoids associated with *Anastrepha fraterculus* (Diptera: Tephritidae) in native fruits: first record of *Aganaspis nordlanderi* (Hymenoptera: Figitidae) in the state of Santa Catarina. **Revista Brasileira de Fruticultura,** Jaboticabal, v. 40, n.3, p.1-5, 2018.

SANTOS, J.P.; LACERDA, A.E.B.; ALVES, A.C.; ANSILIERO, A.A. Índices de infestação da mosca-das-frutas sulamericana em frutíferas nativas em Caçador, SC, Brasil. Actas Portuguesas de Horticultura, Lisboa, v.29, p.86-92, 2018.

SANTOS, J.P. Frutíferas hospedeiras de moscas-das-frutas: arrancálas ou conservá-las? **Agropecuária Catarinense,** Florianópolis, v.35, n.1, p.5-7, 2022.

SANTOS, J.P.; MENEZES-NETTO, A.C.; WERNER, S.S.; LINS JUNIOR, J.C. Infestação de moscas-dasfrutas hospedeiros em nativos na região do Alto Vale do Rio do Peixe, Santa Catarina. Agropecuária Catarinense, Florianópolis, v.35, n.2, p.31-36, 2022.

SILVEIRA NETO, S.S.; MONTEIRO, R.C.; ZUCCHI, R.A.; MORAES, R.C.B. Uso da análise faunística de insetos na avaliação do impacto ambiental. **Scientia Agrícola**, Piracicaba, v.52, n.1, p.9-15, 1995.

ZUCCHI, R.A. Taxonomia. *In*: MALAVASI, A.; ZUCCHI, R.A. (Orgs.). *In*: MALAVASI, A.; ZUCCHI, R.A. (Orgs.). **Moscas-das-frutas de importância econômica no Brasil:** conhecimento básico e aplicado. Ribeirão Preto: Holos, 2000. p.13-24.